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STULV20AA2265-1	Measurement of antivira	al activity of INTERCEPT CU22	FOAM
	INTERCEPT Technology	GmbH	
Spougop	Am Goldberg 2		
SPONSOR	99817 Eisenach		
	GERMANY		
REFERENCE TEST METHOD	ISO 18184:2019 - Measu surfaces	rement of antiviral activity on text	iles and other porous
TEST ITEM			
PRODUCT NAME	INTERCEPT CU22™ FO	AM	
MATRIX OF THE PRODUCT	INTERCEPT CU22 [™] foa Copper	m, Open Cell Foam coated with l	INTERCEPT polymerized
Ватсн	NA	CODE	NA
MANUFACTURING DATE	NA	EXPIRY DATE	NA
MANUFACTURER	INTERCEPT Technology	GmbH	4
ACTIVE INGREDIENT	Polymerized Copper		
PARCEL REGISTRATION N.	IP-LV-2020121-AFW	RECEIVING DATE	April 30 th 2020
STORAGE CONDITIONS	Room temperature		
Analysis Starting Date	May 27 th 2020	ANALYSIS ENDING DATE	June 06 th 2020
EXPERIMENTAL CONDITION	ONS		
TEST TEMPERATURE	Room temperature (25±1	°C) at ≥90%RH	
SPECIMEN DESCRIPTION		coloured PU foam coated with an foam inert uncoated specimen w	Ο,
VIRAL INOCULUM		th known viral titre were applied o was left adsorbing onto the speci	
PRODUCT APPLICATION	NA		
VOLUME APPLIED	NA		
CONTACT TIME	30 minutes, 1 hour, 24 ho	urs (±5 minutes)	
INACTIVATION OF PRODUCT RESIDUES	Dilution-neutralization in o	cell culture medium (no detoxifica	ition needed)
INCUBATION TEMPERATURE	37°C ± 1°C (with 5% CO ₂)		
TEST VIRUS	Bovine Coronavirus (BCo	/) - strain S379 Riems	
CELL LINE	HRT-18 cells (human rec	tal carcinoma cells)	



BioPharma Product Testing

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Check of cytotoxicity of the test item

The test item was not cytotoxic, i.e. its contribution in terms of CPE was not visible in the test.

Assay of viral infectivity (virus titration)

The titre of the starting viral suspension was sufficiently high to at least enable a theoretical viral titre reduction of 4 Log $TCID_{50}$.

Check of viral recovery (untreated surface)

The dose of infectious particles recovered immediately after inoculation (as well as after 30 minutes and 1 hour) from the untreated test specimens was around 6LogTCID_{50} . The dose of infectious particles recovered from each untreated test specimen after contact of 1 h was about 0.3LogTCID_{50} lower than at t_0 . The dose of infectious particles recovered from each untreated test specimen after contact of 24 h was about 1LogTCID_{50} lower than at t_0 . It was 5.00 ± 0.25 LogTCID50/ml.

Check of host cells susceptibility to virus and suppression of antiviral activity (neutralization)

VALIDITY AND EFFICACY CRITERIA

The difference of the average value of $TCID_{50}$ among the cellular cultures treated with the treated samples or untreated samples and then with the viral inoculum and the ones treated only with the viral inoculum (negative control) was $\leq 0.5 \text{ Log}TCID_{50}$.

Accuracy of virus control among the three replicas

The maximum difference of the value of $TCID_{50}$ among the cellular cultures treated with the viral inoculum recovered from the 3 different untreated specimen was ≤ 0.5 Log.

Antiviral efficacy

The LogTCID $_{50}$ reduction factor (R) is calculated as per ISO 18184 :2019 standard, i.e. subtracting the average LogTCID $_{50}$ of treated specimen (At) from the average LogTCID $_{50}$ of untreated specimen (Ut) at the chosen contact times:

The LogTCID₅₀ is calculated by the standard Spearman-Karber method and by the Large Volume Plating method as confirmatory test.

Bovine coronavirus is used as a surrogate virus for SARS-related viruses as it belongs to the same Betacoronavirus genus and showed similar susceptibility to WHO formulations in published studies.



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		Cytotoxicity		
	HRT-18 cell destruction		≤0.50 (Log)	
	Log redu	ctions at the different	contact times	
		30 minutes	1 hour	24 hours
RESULTS	Bovine coronavirus		Average	i.
	(Betacoronavirus 1)	ND	ND	1.19±0.39Log ₁₀
		ND	ND	93%
	See Annex	k N.1 for the detail of t	he test results	
Conclusions	The antiviral treatment causes in the adopted test conditions The treated surface does not h			
Anney	N. 1: PAW DATA ELABODATION			
ANNEX	N. 1: RAW DATA ELABORATION			



Table 1: product description

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Test item [Function]	Product designation	Description	Test surface inoculated and covered by PU-foam
Test squares 2 cm x 2 cm (equipped with the active component(s)	INTERCEPT CU22 Foam	test squares made from PU-foam with a thickness of approx. 2 mm; brown coloured and interspersed with copper-colored metallic-looking particles	2 cm x 2 cm = 4 cm ²
Test squares 2 cm x 2 cm (non-active control)	PU-foam Control	test squares made from PU-foam with a thickness of approx. 1 cm; white coloured with a homogen structure	2 cm x 2 cm = 4 cm ²

Table 2: cytotoxicity control

2 1 1/1	-				Dilution	factor (lg) / VF = 4	1		Titer	Titer				
Product(s)	Exposure	Sample ID	-0,6	-1,2	-1,8	-2,4	-3,0	-3,6	-4,2	per 100 μL (Ig TD _{so})	per 1 mL (lg TD _{so})				
		T-1	0/4 1							≤ 0,30	≤ 1,30				
INTERCEPT CU22 Foam		T-2	0/4							≤ 0,30	≤ 1,30				
	24 h at 25 °C	T-3	0/4							≤ 0,30	≤ 1,30				
	and 90% humidity		1			T-4	0/4							≤ 0,30	≤ 1,30
PU-foam Control		T-5	0/4							≤ 0,30	≤ 1,30				
		T-6	0/4							≤ 0,30	≤ 1,30				

¹ = first number: number of cell cultures with a visible cytotoxic alteration; second number: total number of cell cultures

Table 3: verification of cells susceptibility

	Test sample(s) Sample ID Dilutio						Dilutio	n (lg) /	VF = 4					Titer	
Test sample(s)	Sample ID	factor 1	-0,6	-1,2	-1,8	-2,4	-3,0	-3,6	-4,2	-4,8	-5,4	-6	-6,6	per 100 μL (lg ID ₅₀)	∆ Titer³ (lg ID₅o)
	VK/E-1		4/41	4/4	4/4	4/4	4/4	4/4	3/4	3/4	2/4	0/4		5,1 ± 0,55	-
untreated cells	VK/E-2	n.a.	4/4	4/4	4/4	4/4	4/4	4/4	4/4	4/4	1/4	0/4		5,25 ± 0,30	-
	VK/E-3		4/4	4/4	4/4	4/4	4/4	4/4	3/4	1/4	0/4			4,5 ± 0,42	•
Average vir	rus titer		12/12	12/12	12/12	12/12	12/12	12/12	10/12	8/12	3/12	0/12		4,95 ± 0,27	•
	E-1		4/4	4/4	4/4	4/4	4/4	4/4	2/4	0/4				4,2 ± 0,35	0,75 ± 0,44
INTERCEPT CU22 Foam (treated cells)	E-2	VF = 1	4/4	4/4	4/4	4/4	4/4	3/4	0/4	3/4	1/4	0/4		4,35 ± 0,52	0,60 ± 0,58
	E-3		4/4	4/4	4/4	4/4	4/4	4/4	4/4	2/4	0/4			4,8 ± 0,35	0,15 ± 0,44
Average vir	rus titer		12/12	12/12	12/12	12/12	12/12	11/12	6/12	5/12	1/12	0/12		4,45 ± 0,29	•
	E-4		4/4	4/4	4/4	4/4	4/4	4/4	4/4	3/4	0/4			4,95 ± 0,30	0,0 ± 0,4
PU-foam Control (treated cells)	E-5	VF = 1	4/4	4/4	4/4	4/4	4/4	4/4	4/4	2/4	0/4			4,8 ± 0,35	0,15 ± 0,44
	E-6		4/4	4/4	4/4	4/4	4/4	4/4	3/4	2/4	0/4			4,65 ± 0,46	0,3 ± 0,53
Average vir	Average virus titer			12/12	12/12	12/12	12/12	12/12	11/12	7/12	0/12			4,8 ± 0,20	-

^{1 =} dilution (or dilution factor) of the test sample(s) distributed to the detection cells when the Large Volume Plating (LVP) method was used

² = first number = number of virus positive cells cultures, second number = total number of cell cultures

³ = virus titer A (virus titration on untreated cells) minus virus titer B (virus titration on treated cells)



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Table 4: verification of the suppression of the residual antiviral activity

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Test sample(s)	Sample					Dilutio	n (lg) /	VF = 4					Titer per 100 μL	∆ Titer³	After-effect	Verification acc. clause
rest sample(s)	ID	-0,6	-1,2	-1,8	-2,4	-3,0	-3,6	-4,2	-4,8	-5,4	-6	-6,6	(lg ID ₅₀)	(Ig ID _{so})	present ⁴	10.6 passed 5
	VN-1	4/41	4/4	4/4	4/4	4/4	4/4	4/4	2/4	1/4	1/4	0/4	5,1 ± 0,55	-	-	
Cell culture medium (negative control)	VN-2	4/4	4/4	4/4	4/4	4/4	4/4	4/4	3/4	0/4			4,95 ± 0,30	-	-	n.a.
	VN-3	4/4	4/4	4/4	4/4	4/4	4/4	3/4	1/4	1/4	0/4		4,65 ± 0,52	•	•	n.a.
Average virus titer		12/12	12/12	12/12	12/12	12/12	12/12	11/12	6/12	2/12	1/12	0/12	4,9 ± 0,27		-	
Resuspension medium derived	N-1	4/4	4/4	4/4	4/4	4/4	4/4	4/4	1/4	0/4			4,65 ± 0,30	0,25 ± 0,40	no	
from INTERCEPT CU22 Foam	N-2	4/4	4/4	4/4	4/4	4/4	4/4	4/4	1/4	1/4	0/4		4,8 ± 0,42	0,1 ± 0,50	no	
(treated test specimens)	N-3	4/4	4/4	4/4	4/4	4/4	4/4	3/4	0/4				4,35 ± 0,30	0,55 ± 0,40	no	yes
Average virus titer		12/12	12/12	12/12	12/12	12/12	12/12	11/12	2/12	1/12	0/12		4,6 ± 0,20	antiviral	specimen	Δ Titer of reference specimen
Decree and a second second second	N-4	4/4	4/4	4/4	4/4	4/4	4/4	3/4	2/4	1/4	1/4	0/4	4,95 ± 0,62	-0,05 ± 0,68	no	minus antiviral specimen =
Resuspension medium derived from PU-foam Control	N-5	4/4	4/4	4/4	4/4	4/4	4/4	4/4	4/4	0/4			5,1 ± 0,0	-0,2 ± 0,27	no	0,25 ± 0,33
(untreated test specimens)	N-6	4/4	4/4	4/4	4/4	4/4	4/4	3/4	1/4	0/4			4,5 ± 0,42	0,4 ± 0,5	no	
Average virus titer		12/12	12/12	12/12	12/12	12/12	12/12	10/12	7/12	1/12	1/12	0/12	4,85 ± 0,26	reference	specimen	

¹ = the test virus was added directly into the test sample as resuspended from the test item. Incubation was t = 30 min. at 25 °C.
² = first number = number of virus positive cells cultures, second number = total number of cell cultures

Table 5a: titration of the virus suspension and the virus material recovered from the virus control

						Dilutio	n (lg) /	VF = 4					Titer per	Ø Titer per	Verification
Test sample(s)	Sample ID	-0,6	-1,2	-1,8	-2,4	-3,0	-3,6	-4,2	-4,8	-5,4	-6	-6,6	100 μL (lg ID ₅₀)	1 mL (lg ID ₅₀)	passed ²
	Aus-1	4/41	4/4	4/4	4/4	4/4	4/4	3/4	3/4	1/4	1/4	0/4	5,1 ± 0,60		
Virus suspension when added directly to the	Aus-2	4/4	4/4	4/4	4/4	4/4	4/4	3/4	2/4	0/4			4,65 ± 0,46	5,90 ± 0,26	yes Virus titer of stock
resuspension medium	Aus-3	4/4	4/4	4/4	4/4	4/4	4/4	4/4	3/4	0/4			4,95 ± 0,30		virus suspension: IgID ₅₀ = 7,59/mL
Average virus titer		12/12	12/12	12/12	12/12	12/12	12/12	10/12	8/12	1/12	1/12	0/12	4,9 ± 0,26		
Virus material as recovered from	VK-1	4/4	4/4	4/4	4/4	4/4	4/4	4/4	3/4	0/4			4,95 ± 0,30		
the non-coated control test item after t = 0 min.	VK-2	4/4	4/4	4/4	4/4	4/4	4/4	4/4	4/4	0/4	1/4	0/4	5,25 ± 0,30	6.1 ± 0.24	
arter t = 0 min.	VK-3	4/4	4/4	4/4	4/4	4/4	4/4	4/4	4/4	0/4			5,1 ± 0,0	0,1 ± 0,24	
Average virus titer		12/12	12/12	12/12	12/12	12/12	12/12	12/12	11/12	0/12	1/12	0/12	5,1 ± 0,14		yes Δ Titer of (t = 0)
Management of the second of th	VK-4	4/4	4/4	4/4	4/4	4/4	3/4	1/4	1/4	0/4			4,05 ± 0,52		minus (t = 24) = 1,1 ± 0,35
Virus material as recovered from the non-coated control test item after t = 24 hours	VK-5	4/4	4/4	4/4	4/4	4/4	4/4	0/4					3,9 ± 0,0	50.005	
arter t = 24 nours	VK-6	4/4	4/4	4/4	4/4	4/4	2/4	3/4	0/4				4,05 ± 0,46	5,0 ± 0,25	
Average virus titer		12/12	12/12	12/12	12/12	12/12	9/12	4/12	1/12	0/12			4,0 ± 0,25		

^{1 =} first number = number of virus positive cells cultures, second number = total number of cell cultures

^{3 =} virus titer A (negative control [VN]) minus virus titer B (treated test specimen [N-1 to N-3]) or minus virus titer c (untreated test specimen [N-4 to N-6])

^{4 =} according to the EN 14476 an ongoing residual disinfecting activity (after effect) of the product(s) applies as not given when Δ Titer is ≤ $\lg 0.5$

 $^{^{\}text{5}}$ = verification acc. ISO 18184, clause 10.6.2 is passed when Δ Titer is $\,\leq$ lg 0,5



Table 5b: titer of virus control at 30 minutes and 1 hour

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T	6					Dilutio	on (lg) /	VF = 4					Titer per	Ø Titer per	Average virus titer
Test sample(s)	Sample ID	-0,6	-1,2	-1,8	-2,4	-3,0	-3,6	-4,2	-4,8	-5,4	-6	-6,6	100 μL (lg ID ₅₀)	1 mL (lg ID _{so})	
Virus material as recovered from the non-coated control test item	VK-7	4/41	4/4	4/4	4/4	4/4	4/4	3/4	2/4	0/4			4,65 ± 0,46		
after t = 30 min.	VK-8	4/4	4/4	4/4	4/4	4/4	4/4	4/4	1/4	0/4			4,65 ± 0,30	5,65 ± 0,27	
Average virus titer		8/8	8/8	8/8	8/8	8/8	8/8	7/8	3/8	0/8			4,65 ± 0,27		F 70 : 0 40
Virus material as recovered from	VK-9	4/4	4/4	4/4	4/4	4/4	4/4	4/4	0/4				4,5 ± 0,0	5,8 ± 0,27	5,73 ± 0,18
the non-coated control test item after t = 1 hour	VK-10	4/4	4/4	4/4	4/4	4/4	4/4	4/4	3/4	1/4	0/4		5,1 ± 0,42	Δ Titer of (t = 0) minus (t = 1) =	
Average virus titer		8/8	8/8	8/8	8/8	8/8	8/8	8/8	3/8	1/8	0/8		4,8 ± 0,27	0,3 ± 0,36	

^{1 =} first number = number of virus positive cells cultures, second number = total number of cell cultures

Table 6: Maximum RF (reduction factors) at 30 minutes, 1 hour and 24 hours

Test virus	Dilution ¹ factor	Incubation time (h)	Virus titer per 1 mL ¹ [Ig ID∞ ± KI∞ x]	detection limit [lg ID ₅₀ / mL]	max. detectable virus reduction (RF _{max}) ²
			Virus titration using the limiting dilu	ution method (Spearman & Kärber)	
		24	5,0 ± 0,25	Ig ID ₅₀ = 1,30	3,7
	VF = 4	1	5,73 ± 0,18	Ig ID ₅₀ = 1,30	4.43
Bovine Coronavirus		0,5	5,75 ± 0,16	ig iD ₅₀ = 1,50	4,43
(S379 Riems)		٧	irus titration by Large Volume Platir	ng (LVP) inoculating 48 cell cultures	3
		24	5,0 ± 0,25	Ig ID ₅₀ = -0,83	5,83
	VF = 1	1	F 72 + 0.19	Ig ID ₅₀ = -0,83	6.56
		0,5	5,73 ± 0,18	ig i∪ ₅₀ = -∪,83	6,56

⁼ input virus (virus control), cf. Tab. 5

² = maximum detectable virus reduction (RF_{max}) when no residual virus was detectable. With LVP the detection limit was calculated with the modified Poisson-Formula (cf. Ref 7).

 $^{^{3}}$ = when 48 cell culture units were inoculated; V = 10,2 mL and v = 9,6 mL.



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Table 7: inactivation tests (3 test replicas) by Spearman-Karber method (S-K)

Table 1. Illactivation tests (o lest replica	eplicas) by Spearman-Karber method (S-K)													
Test sample(s)	Sample ID	Incubation					Dilutio	n (lg) /	VF = 4					Titer per	Ø Titer
rest sample(s)	Sample 1D	time	-0,6	-1,2	-1,8	-2,4	-3,0	-3,6	-4,2	-4,8	-5,4	-6	-6,6	100 μL (lg ID ₅₀)	per 1 mL (lg ID ₅₀)
	In-1		4/4 1	4/4	4/4	4/4	2/4	2/4	1/4	0/4				3,45 ± 0,5	
INTERCEPT CU22 Foam	In-2	24.5	4/4	4/4	4/4	3/4	1/4	0/4						2,7 ± 0,37	3,8 ± 0,30
	In-3	24 h	4/4	4/4	3/4	2/4	0/4							2,25 ± 0,4	3,610,30
Average virus tite	er		12/12	12/12	11/12	9/12	3/12	2/12	1/12	0/12				2,8 ± 0,30	
	In-4		4/4	4/4	4/4	4/4	4/4	4/4	4/4	3/4	2/4	0/4		5,25 ± 0,46	
INTERCEPT CU22 Foam	In-5	0,5 h	4/4	4/4	4/4	4/4	4/4	4/4	4/4	1/4	3/4	1/4	0/4	5,25 ± 0,52	6,2 ± 0,29
	In-6	0,5 H	4/4	4/4	4/4	4/4	4/4	4/4	3/4	4/4	1/4	0/4		5,1 ± 0,42	0,210,29
Average virus tite	er		12/12	12/12	12/12	12/12	12/12	12/12	11/12	8/12	6/12	1/12	0/12	5,2 ± 0,29	
	In-7		4/4	4/4	4/4	4/4	4/4	4/4	4/4	3/4	0/4			4,95 ± 0,30	
INTERCEPT CU22 Foam	In-8		4/4	4/4	4/4	4/4	4/4	4/4	4/4	1/4	0/4	1/4	0/4	4,8 ± 0,42	E 0E + 0 30
	In-9	1 h	4/4	4/4	4/4	4/4	4/4	4/4	4/4	4/4	0/4	0/4		5,1 ± 0,0	5,95 ± 0,20
Average virus tite	Average virus titer		12/12	12/12	12/12	12/12	12/12	12/12	12/12	8/12	0/12	1/12	0/12	4,95 ± 0,20	

^{1 =} first number = number of virus positive cells cultures, second number = total number of cell cultures.

Table 8: estimation of the reduction factors (RF)

Table 8: estimation of the redu	Clion ractors (R	<i>「</i>)					
Droduct(s)	Samula ID	Incubation	lg ID ₅₀ /mL [l	g ID ₅₀ ± KI _{95%}]	Reduction fa	ctor [± Kl _{95%}]	
Product(s)	Sample ID	time	Virus input 1	Residual virus ²	Virus reduction	Average ³	
	In-1			4,45 ± 0,5	0,55 ± 0,56	1,19 ± 0,39	
	In-2	24 h	5,0 ± 0,25	3,7 ± 0,37	1,3 ± 0,45	(equivalent to an average reduction	
	In-3			3,25 ± 0,4	1,75 ± 0,47	of 93% within 24 h)	
	In-4			6,25 ± 0,46	-0,52 ± 0,49		
INTERCEPT CU22 Foam	In-5	0,5 h	5,73 ± 0,18	6,25 ± 0,52	-0,52 ± 0,55	Not detectable	
	In-6			6,1 ± 0,42	-0,37 ± 0,46		
	In-7			5,95 ± 0,30	-0,22 ± 0,35		
	In-8	1 h	5,73 ± 0,18	5,8 ± 0,42	-0,07 ± 0,46	Not detectable	
	In-9			6,1 ± 0,0	-0,37 ± 0,18		

^{1 =} amount of input virus (virus control; cf. Tab. 5)

^{2 =} amount of residual virus with respect to the cytotoxicity titer (cf. Tab. 2)

 $^{^{3}}$ = titer of input virus (lg ID₅₀) after t = 24 h (U_t) minus titer of residual virus (lg ID₅₀) after t = 24 h (A_t) [R = U_t - A_t]



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Inactivation of the BCoV by INTERCEPT CU22TM Foam at 25°C within 24 hours

Antiviral validation using the quantitative carrier test according to ISO 21702:2019

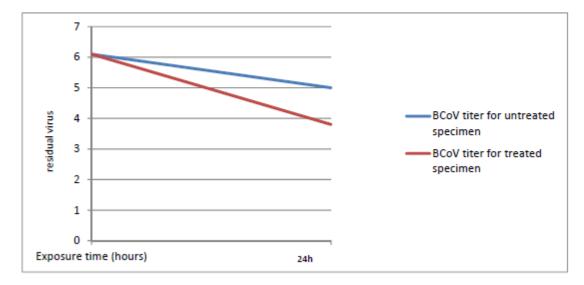


Fig.1

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The test results relate only to the tested items. Sampling, except specific indication on test report, is always intended to be made by the Sponsor. Characterization of the test sample is under Sponsor responsibility.